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IN-VITRO EVALUATION OF ANTIOXIDANT ACTIVITY OF ALBIZIA LEBBECK BARK

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ABSTRACT

The present study was designed to evaluate the *in-vitro* antioxidant potential of methanolic extract and isolated compound from *Albizia lebeck* bark. The isolation of compound from methanolic extract was carried out by column chromatography technique and purity of isolated compound was checked by thin layer chromatography. The total phenolic contents of methanolic extract and isolated compound were determined by using Folin - Ciocalteu method. The total phenolic content was high in the isolated compound (84.44 mg/g gallic acid equivalent GAE) as compared to methanolic extract (80.25 mg/g gallic acid equivalent GAE). The methanolic extract and isolated compound were investigated for free radical scavenging activity of the 1, 1-diphenyl-2-picrylhydrazyl radical (DPPH) and reducing power assay. The results of DPPH free radical scavenging at 1000 µg/ml indicated maximum antioxidant activity of 91.82% and 90.08% respectively. The reducing power of isolated compound and methanol extract were found to be 2.799 nm and 2.2148 nm respectively. The phytochemical screening revealed the presence of alkaloids and polyphenolic compounds. This suggests a potential utility of the plant as a source of phenolic antioxidants and may provide leads in the ongoing research for natural antioxidants from Indian medicinal plants to be used in treating diseases related to free radical reactions.

Key Words:- *Albizia lebeck*, Free radical scavenging activity, Antioxidant activity.

INTRODUCTION

Antioxidants are believed to quench free radicals. Free radicals are atoms or molecules with a single unpaired electron which makes them highly reactive. Oxidative free radicals are generated by metabolic reactions. Free radicals create a chain reaction leading to membrane lipid peroxidation, DNA damage etc. Free radical oxidation has been implicated in Cancer, atherosclerosis, neurodegenerative diseases and inflammatory bowel disease (Tripathi, 2008). Antioxidants are added to pharmaceutical formulations as redox systems possessing higher oxidative potential than the drug that they are designed to protect, or as chain inhibitors of radical

induced decomposition. In general, the effect of oxidants is to break up the chain formed during a hydrogen atom or an electron to free radical receiving the excess energy possessed by the active molecule (Lachman, 1986).

Albizia lebeck Linn. (Mimosaceae) commonly called as *siris* is widely used in the treatment of many ailments. It has widely distributed all over India, mostly in Maharashtra, Punjab, Gujarat, Karnataka and Madhya Pradesh. The bark of *Albizia lebeck* contains tannins of condensed type, isomers of leucocyanidin, melacacidine, new leucoantho-cyanidin and lebecacidin. It also contains triedelin and t_3 -sitosterol. The leaves of *Albizia lebeck* used as an antiseptic and wound healing property. The bark of *Albizia lebeck* used in the treatment of piles. It also possesses anti-spermatogenic and anti-inflammatory activity (Khare, 2007; Kirtikar, 1999). This study was aimed to investigate antioxidant potential of

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Albizialebeck bark. Therefore an effort is made to contribute to establish scientific evidence in this regard.

MATERIALS

Plant material

The bark of *Albizialebeck* was collected from the Botanical garden, Nagpur and taxonomically authenticated from the Department of Botany, Rashtrasant Tukadoji Maharaj Nagpur University, Nagpur. A Voucher specimen (No. 9588) has been deposited in the Herbarium of the same department.

EXPERIMENTAL AND RESULTS

Preparation of extract

Bark of *Albizialebeck* was dried in shade and powdered in hand grinder to get coarse powder. The powdered bark was extracted for 30 h in a Soxhlet apparatus using methanol. The methanolic extract was concentrated at low pressure by distillation and finally air dried.

Preliminary phytochemical screening

The plants may contain phytochemical constituents like alkaloids, glycosides, sterols, tannins, saponins, sugars, etc. These compounds are secondary metabolites mainly responsible for their physiological and therapeutic effects. The methanol extract and isolated compound from *Albizialebeck* bark were subjected to preliminary phytochemical screening for the detection of various plant constituents (Lala, 1985). The results of preliminary phytochemical screening are given in Table 1.

Methanolic extract showed positive test for alkaloids which are targeted bioactive constituents hence it was selected for isolation of active constituent. The isolation procedure was carried out by column chromatography method and purity was checked by thin layer chromatography method. The results of thin layer chromatography of methanol extract are shown in Table 2. Mobile phase with Butanol: Glacial Acetic acid: Water (8:1:1) showed maximum resolution and reproductive results. In iodine vapour and after spraying with 50% H_2SO_4 showed 4 spots having R_f values 0.31, 0.49, 0.53 and 0.65.

Column chromatography of methanolic extract

Preparation of column

The glass column of about 1.2 m length and 3 cm in diameter was used. The column was cleaned thoroughly using chromic acid and washed with distilled water. Silica gel 60-120 mesh was activated at 120° in hot air oven for 1 h and mixed with petroleum ether to form slurry. The column was fixed vertically on a stand and a cotton plug was inserted to bottom of the column. The column was

filled with petroleum ether and the slurry of silica gel was poured slowly from the top. The solvent used for elution was maintained 10 cm above the cotton plug. The methanolic extract (40 g) was triturated with small quantity of silica gel and poured from top of the column. A plug of cotton was placed over it and solvent is maintained 10 cm above the cotton plug (Stahl, 2005; Silverstein, 1991).

Elution of column

The column was eluted successively with different solvents (from nonpolar to polar) and mixture of solvents, in increasing order of polarity and 30 ml of fractions were collected every time. Eluted fractions were subjected to thin layer chromatography for every solvents or solvent mixture. For homogeneity test of each fraction thin layer chromatogram was run by using different solvent systems and 50% sulphuric acid was used as a detecting reagent. The results of thin layer chromatography of eluted fractions and isolated compound are shown in table 3 and 4 respectively. The result of identification of isolated compound is shown in table 5. The fractions having same TLC pattern were mixed to form single fraction.

Interpretation of isolated compound

Isolated compound gives positive test for alkaloid which indicate, the compound have alkaloid in nature. It is soluble in methanol having melting point 144-146°. Thin layer chromatography of isolated compound showed R_f value 0.65 using mobile phase butanol: acetic acid: water (6:1:3). UV spectra of isolated compound showed λ_{max} at 430 nm. IR spectra of isolated compound also support the alkaloidal nature of the isolated compound. Bands at 3550 cm^{-1} , 2931 cm^{-1} and 1639 cm^{-1} indicates N-H, C-H and C=O stretching of the compound. PMR spectra of isolated compound also suggest peak at 7.27 ppm singlet indicates that the isolated compound is indole alkaloid in nature. Other values of PMR spectra also support the indole alkaloidal nature of the compound. Mass spectra of isolated compound showed molecular ion peak at m/e 336 which indicates molecular weight of the isolated compound was found to be 336.

Quantitative estimation of total phenolic contents (folin – ciocalteu method)

Standard Gallic acid was accurately weighed in volumetric flask and dissolved in 100 ml of distilled water and various concentrations of gallic acid (0.2-1.0 mg/ml) were prepared. From these various concentrations of gallic acid, 1ml was mixed with 5 ml of Folin- ciocalteu reagent (diluted tenfold) and then 4ml of (7.5%) sodium carbonate. The absorption was measured after 30 m at 20°

at 765 nm and the calibration curve was drawn. Methanolic extract (1 ml) was mixed with the same reagents as shown above, and after 30 min the absorption was measured at 765 nm for the determination of phenolic contents. Total phenolic content (%) in methanolic extract and isolated compound were calculated as (GAE) Gallic acid equivalent (Miniauskas, 2004). Standard calibration curve of gallic acid is shown in table 6 and figure 6. The results for total phenolic content in methanolic extract and isolated compound are shown in table 7.

$$\text{GAE} = [(C \times V) / M] \times 100$$

Where,

C = the conc. of Gallic acid established from calibration curve (mg/ml)

V = Volume of extract (ml)

M = the weight of dried plant extract (mg)

Evaluation of *in-vitro* antioxidant activity

DPPH assay

Methanolic solution of isolated compound and methanolic extract were prepared at various concentrations (100, 200, 400, 800 and 1000 µg/ml). To a set of test tubes, 2.9 ml of DPPH solution (100 µg/ml in methanol) and 0.1 ml of varying concentrations of test sample were added. The mixture was then shaken vigorously and allowed it to stand in dark for 30 m and absorbance was measured using a UV spectrophotometer at 517 nm (Miliauskas, 2001). A control solution was consisting of

0.1 ml of methanol and 2.9 ml of DPPH radical solution (Desai and Wadekar, 2008). Percentage scavenging of DPPH radical was calculated by comparing the absorbance between the test, mixture and control.

Percentage scavenging of DPPH radical was shown in table 8 and figure 2.

$$\% \text{ Scavenging of DPPH} = \frac{\text{Absorbance (control)} - \text{Absorbance (sample)}}{\text{Absorbance (Control)}} \times 100$$

Reducing power method

The reducing power of the extracts was carried out according to the conventional method. Various concentrations of methanolic extract and isolated compound (100, 200, 400, 800, 1000 µg/ml) in 1.0 ml of deionized water were mixed with phosphate buffer (2.5 ml, 0.2M, pH 6.6) and 1% of potassium ferricyanide (2.5 ml). The mixture was incubated at 50° for 20 m and aliquots of trichloroacetic acid (2.5 ml, 10%) were added to the mixture, which was centrifuged at 1036 x g for 10 min. The upper layer of solution (2.5 ml) was mixed with distilled water (2.5 ml) and a freshly prepared FeCl₃ solution (0.5 ml, 1%). The absorbance was measured at 700 nm (Scherer and Godoy, 2009; Gulcin et al., 2004). Increased absorbance of the reaction mixture indicated increased reducing power. The results of reducing power of methanolic extract, isolated compound and ascorbic acid are shown in table 9 and figure 3.

Table1. Preliminary phytochemical screening of *Albizialebeck*

Plant Constituents	Tests	Methanolic extract	Isolated compound
Test for Sterols	Liebermann's test	--	--
	Salkowaski Reaction	--	--
	Liebermann-Burchard Reaction	--	--
Test for Alkaloids	Dragendorff's Reagent	++	++
	Mayer's Reagent	++	++
	Wagner's Reagent	++	++
	Hager's Reagent	--	--
	Tannic acid	++	++
Test for Glycoside	Killer-Killani test	++	++
	Legal test	++	++
	Baljet test	++	++
Test for Flavonoids	Shinoda test	--	--
Test for Saponin	Foam test	++	++
	Heamolysis test	++	++
Test for Tannins	5% FeCl ₃	++	++
Test for Protiens	Biuret test	++	++
	Million's test	++	++

-- Absent, + Present, ++ Present in higher concentration

Table 2. Thin layer chromatography of methanolic extract of *Albizialebeck*

Sample spot	Mobile phase used	No. of spots obtained with different locating agent			R _f value
		UV	Iodine vapour	50% H ₂ SO ₄	
Methanolic extract	Methanol	2	3	2	0.17, 0.23
	Ethyl acetate: chloroform(3:7)	2	3	3	0.34, 0.55, 0.67
	Butanol: Glacial acetic acid: Water (8:1:1)	3	4	4	0.31, 0.49, 0.53, 0.65

Table 3. Thin layer chromatography of eluted fractions

Mobile phase used for TLC	Fraction Number	Iodine vapours	
		No. of Spots	R _f
Petroleum ether	1-12	0	0.00
Benzene	13-18	0	0.00
Benzene: Ethyl acetate (9:1)	19-27	1	0.51
Benzene: Ethyl acetate (8:2)	28-48	1	0.58
Benzene: Ethyl acetate (8:2)	49-68	1	0.62
Benzene: Ethyl acetate(7:3)	69-87	1	0.73
Ethyl acetate	88-101	2	0.76, 0.80
Ethyl acetate: Acetone (9:1)	102-112	2	0.78, 0.80
Ethyl acetate: Acetone (8:2)	113-122	2	0.79, 0.82
Butanol: acetic acid: water (6:1:3)	123-148	1	0.65
Butanol: acetic acid: water (6:1:3)	149-194	1	0.65
Butanol: acetic acid: water (6:1:3)	195-220	1	0.65
Butanol: acetic acid: water (6:1:3)	221-240	1	0.65
Butanol: acetic acid: water (6:1:3)	241-267	1	0.65

Table 4. Thin layer chromatography of isolated compound

Sample spot applied	Mobile phase used	No. of spot obtained with different locating agent		R _f value
		Iodine vapour	Dragendorffs reagent	
Isolated compound	Butanol: acetic acid: water (6:1:3)	1	1	0.65

Table 5: Identification of isolated compound

Parameter	Isolated compound (Alkaloid)
Colour	Cremish brown
Solubility	Methanol
Appearance	Fine powder
Melting Point	144-146°
Chromogenic reagent	Iodine vapour, Dragendorff's reagent
R _f value	0.65

Table 6. Standard calibration curve of gallic acid

Sr. No.	Concentration (mg/ml)	Absorbance (765 nm)
1	0.2	0.2140
2	0.4	0.3998
3	0.6	0.5001
4	0.8	0.6101
5	1.0	0.8044

Table 7. Total phenolic content in methanolic extract and isolated compound

Sr.No.	Sample	Absorption at 765nm	GAE
1	Methanolic extract	0.6543	80.25
2	Isolated compound	0.6832	84.44

Table 8. Results of percentage scavenging activity

Sr. No	Samples	Concentration $\mu\text{g/ml}$ and % scavenging of DPPH				
		100	200	400	800	1000
1.	Methanolic extract	20.04	32.63	62.15	84.14	90.08
2.	Isolated compound	22.51	34.95	67.59	86.40	91.82
3.	Ascorbic acid	24.96	37.13	71.21	88.09	95.44

Table9.Reducing power of methanolic extract, isolated compound and ascorbic acid

Sr. No.	Sample	Concentration $\mu\text{g/ml}$ and Absorbance at 700 nm				
		100	200	400	800	1000
1.	Methanolic extract	0.4899	0.8702	1.3114	2.0998	2.2148
4.	Isolated compound	0.6701	0.9065	1.7236	2.2054	2.7992
5.	Ascorbic acid	1.088	1.508	2.205	3.612	3.981

Spectral data of isolated compound

UV spectra: λ max at 430 nm.

IR: ν^{KBr} max (cm^{-1}): 3350, 2931, 2360, 2339, 1606, 1639, 1516, 1447, 1360, 1130, 1067, 860, 858, 830, 777, 608.

^1H NMR: PMR δ ppm-7.27(s), 6.82, 6.79(d), 6.73(s), 6.69, 6.62(d), 6.02, 5.98(d), 4.19(s), 3.74, 3.69(d), 3.30(s), 3.05, 3.03, 3.02(t), 2.79, 2.77, 2.76(t), 2.48(s).

^{13}C NMR: δ ppm-150, 147, 146, 144, 141, 129, 128, 124, 122, 119, 112, 108, 104, 101, 95, 60, 56, 49, 48, 40, 39, 29.

Mass spectra: Molecular ion peak were observed at $M+1 - 337$ and other fragments at m/e 336, 330, 325, 320, 292, 278, 255, 240, 220.

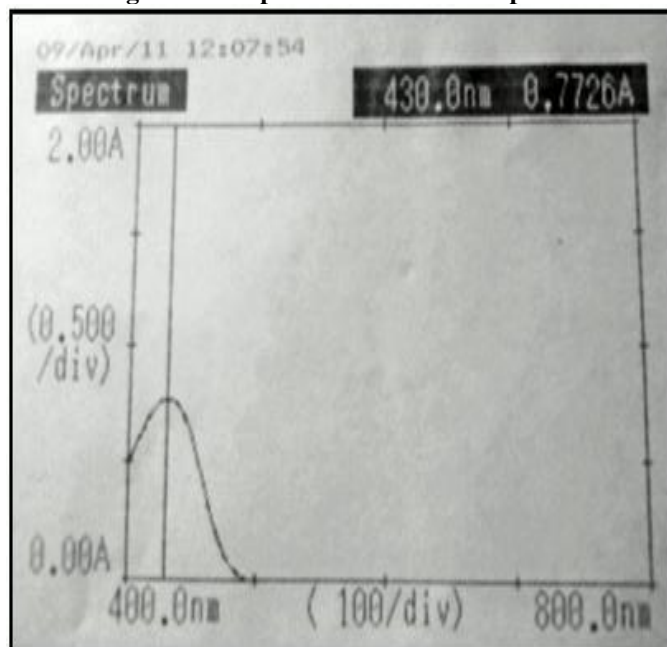
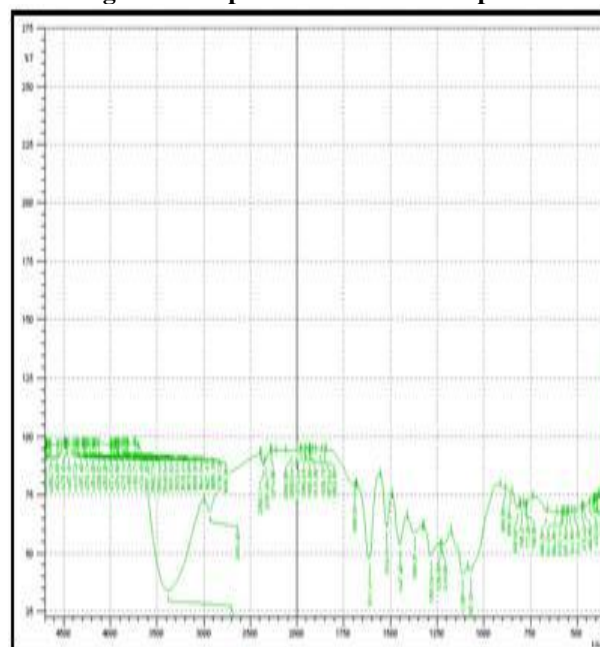
Figure1. UV spectra of isolated compound**Figure 2.IR spectra of isolated compound**

Figure 3. ¹H NMR spectra of isolated compound

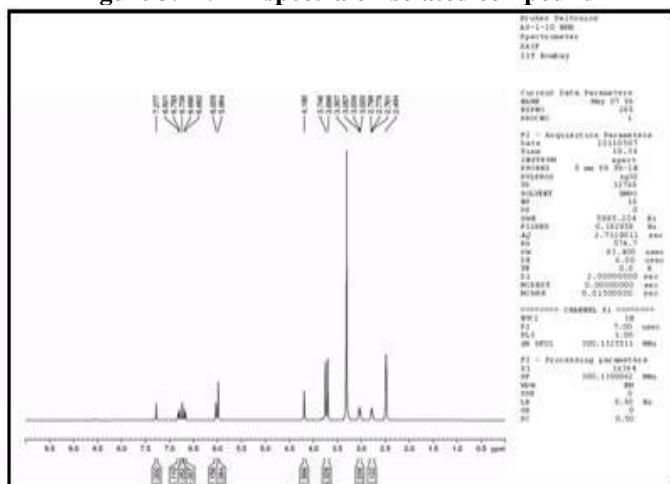


Figure 4. ¹³C NMR spectra of isolated compound

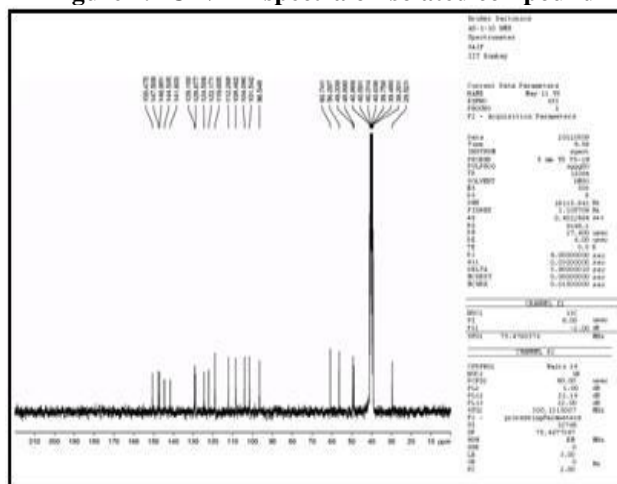


Figure 5. Mass spectra of isolated compound

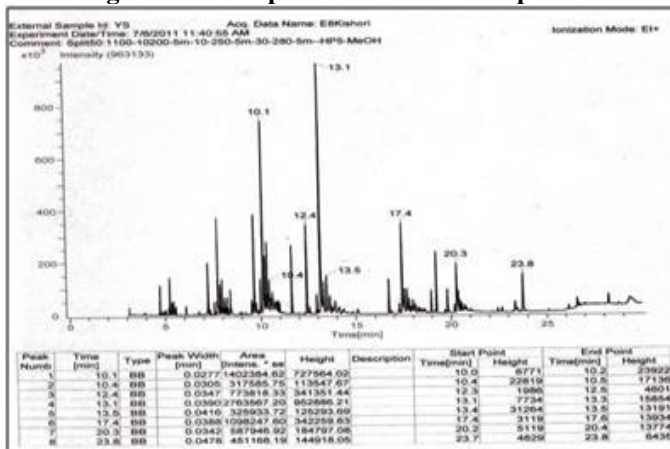


Figure 6. Standard calibration curve of Gallic acid

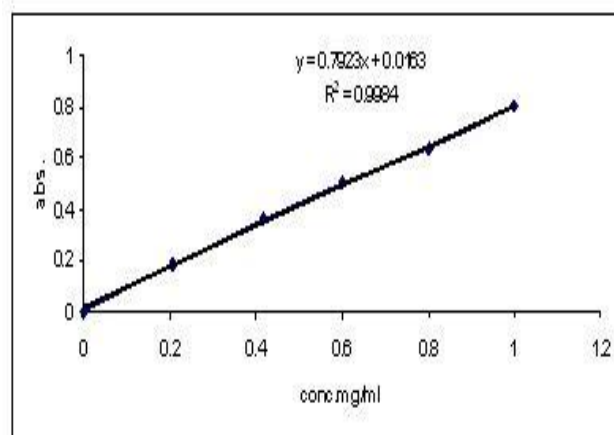


Figure 7. Percentage scavenging activity of DPPH

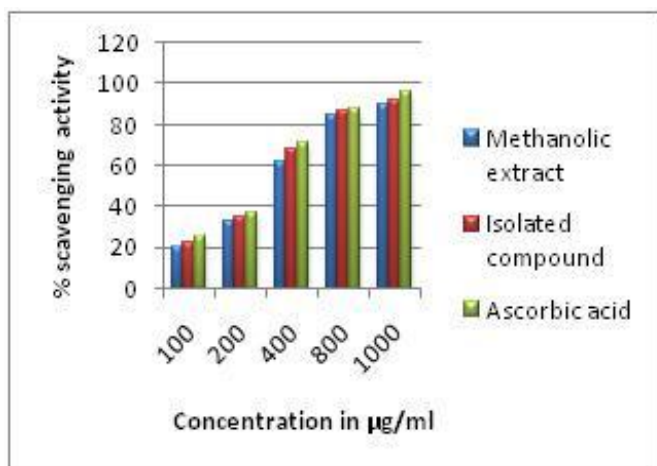
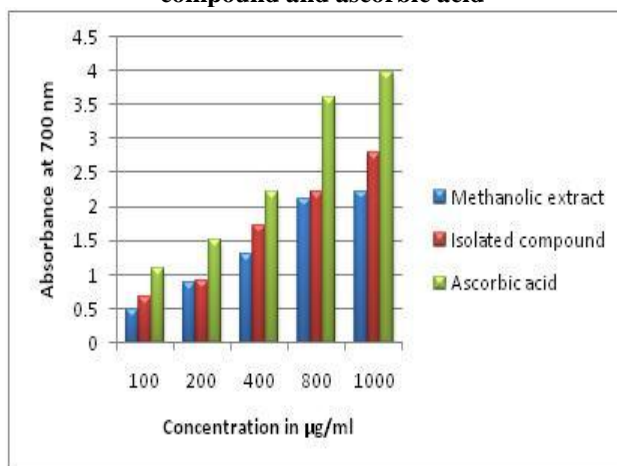


Figure 8. Reducing power of methanolic extract, isolated compound and ascorbic acid



DISCUSSION AND CONCLUSION

The present work deals with the study of extraction of *Albizialebeck*, its evaluation and validation for anti-proliferative activity. Bark was extracted with different organic solvents with increasing order of polarity namely petroleum ether, methanol and hydro alcoholic solution, respective extracts were obtained. Preliminary phytochemical screening showed that methanolic extract contain alkaloids and due to which extract was selected for isolation. Isolation of methanolic extract was carried out by column chromatography. Purity of isolated compound was checked by single spot on TLC. Isolated compound gave positive test for alkaloid (Dragendorffs test), indicating that isolated compound is alkaloid in nature. Isolated compound was screened for antioxidant activity by performing *in vitro* assay method namely DPPH radical scavenging and reducing power method. Isolated compound showed higher antioxidant activity because it contains higher concentration of polyphenolic compound. The present study was designed to evaluate the *in-vitro* antioxidant potential of methanolic extract and isolated compound from *Albizialebeck* bark. The isolation of compound from methanolic extract was carried out by

column chromatography technique and purity of isolated compound was checked by thin layer chromatography. The total phenolic contents of methanolic extract and isolated compound were determined by using Folin - Ciocalteu method. The total phenolic content was high in the isolated compound (84.44 mg/g gallic acid equivalent GAE) as compared to methanolic extract (80.25 mg/g gallic acid equivalent GAE). The methanolic extract and isolated compound were investigated for free radical scavenging activity of the 1, 1-diphenyl-2-picrylhydrazyl radical (DPPH) and reducing power assay. The results of DPPH free radical scavenging at 1000 µg/ml indicated maximum antioxidant activity of 91.82% and 90.08% respectively. The reducing power of isolated compound and methanol extract were found to be 2.799 nm and 2.2148 nm respectively. The phytochemical screening revealed the presence of alkaloids and polyphenolic compounds. This suggest a potential utility of the plant as a source of phenolic antioxidants and may provide leads in the ongoing research for natural antioxidants from Indian medicinal plants to be used in treating diseases related to free radical reactions.

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